

ENDOGENOUS CONSTITUTIVELY ACTIVATED G PROTEIN-COUPLED ORPHAN RECEPTORS

The benefit of commonly owned: (1) Provisional Patent Application Serial Number 60/094879, filed July 31, 1998; (2) Provisional Patent Application Serial Number 60/106300, filed October 30, 1998; (3) Provisional Patent Application Serial Number 60/110,906, filed December 4, 1998, and (4) Provisional Application Serial Number 60/121,851, filed February 26, 1999 is hereby claimed. This patent document is related to U.S. Serial Number 09/060,188, filed April 14, 1998. The entire disclosures of each of the foregoing patent applications are incorporated herein by reference.

FIELD OF THE INVENTION

The invention disclosed in this patent document relates to transmembrane receptors, more particularly to endogenous, constitutively active G protein-coupled receptors for which the endogenous ligand is unknown, and most particularly to the use of such receptors for the direct identification of candidate compounds via screening as agonists, partial agonists or inverse agonists to such receptors.

BACKGROUND OF THE INVENTION

A. G protein-coupled receptors

G protein-coupled receptors share a common structural motif. All these receptors have seven sequences of between 22 to 24 hydrophobic amino acids that form seven alpha helices, each of which spans the membrane. The transmembrane helices are joined by strands of amino acids having a larger loop between the fourth and fifth transmembrane helix on the extracellular side of the membrane. Another larger loop, composed primarily of hydrophilic amino acids, joins transmembrane helices five and six on the intracellular side of the membrane. The carboxy terminus of the receptor lies intracellularly with the amino terminus in the extracellular space. It is thought that the loop joining helices five and six, as well as the carboxy terminus, interact with the G protein. Currently, Gq, Gs, Gi, and Go are G proteins that have been identified. The general structure of G protein-coupled receptors is shown in Figure 1.

Under physiological conditions, G protein-coupled receptors exist in the cell membrane in equilibrium between two different states or conformations: an "inactive" state and an "active" state. As shown schematically in Figure 2, a receptor in an inactive state is unable to link to the intracellular transduction pathway to produce a biological response. Changing the receptor conformation to the active state allows linkage to the transduction pathway and produces a biological response.

A receptor may be stabilized in an active state by an endogenous ligand or an exogenous agonist ligand. Recent discoveries such as, including but not exclusively limited to, modifications to the amino acid sequence of the receptor provide means other than ligands to stabilize the active

state conformation. These means effectively stabilize the receptor in an active state by simulating the effect of a ligand binding to the receptor. Stabilization by such ligand-independent means is termed "constitutive receptor activation." A receptor for which the endogenous ligand is unknown or not identified is referred to as an "orphan receptor."

B. Traditional Compound Screening

Generally, the use of an orphan receptor for screening purposes to identify compounds that modulate a biological response associated with such receptor has not been possible. This is because the traditional "dogma" regarding screening of compounds mandates that the ligand for the receptor be known, whereby compounds that competitively bind with the receptor, *i.e.*, by interfering or blocking the binding of the natural ligand with the receptor, are selected. By definition, then, this approach has no applicability with respect to orphan receptors. Thus, by adhering to this dogmatic approach to the discovery of therapeutics, the art, in essence, has taught and has been taught to forsake the use of orphan receptors unless and until the natural ligand for the receptor is discovered. The pursuit of an endogenous ligand for an orphan receptor can take several years and cost millions of dollars.

Furthermore, and given that there are an estimated 2,000 G protein-coupled receptors in the human genome, the majority of which being orphan receptors, the traditional dogma castigates a creative approach to the discovery of therapeutics to these receptors.

C. Exemplary Orphan Receptors: GPR3, GPR4, GPR6, GPR12, GPR21, GHSR, OGR1 and AL022171

GPR3 is a 330 amino acid G protein coupled receptor for which the endogenous ligand is unknown. (Marchese, A. et al. (1994) *Genomics* 23:609; *see also*, Iismaa, T.P. et al (1994) *Genomics* 24:391; *see* Figure 1 for reported nucleic acid and amino acid sequence.) GPR3 is constitutively active in its endogenous form. (Eggerick, D. et al. (1995) *Biochem. J.* 389:837).

GPR12 is a 334 amino acid homolog of GPR3; the endogenous ligand for GPR12 is unknown (Song, Z. -H., et al (1995) *Genomics*, 28:347; *see* Figure 1 for reported amino acid sequence). GPR6 is a 362 amino acid homolog of GPR3; the endogenous ligand for GPR6 is unknown (Song, Z.-H. et al, *supra.*; *see* Figure 1 for reported amino acid sequence). GPR6 transcripts are reported to be abundant in the human putamen and to a lesser extent in the frontal cortex, hippocampus, and hypothalamus (Heiber, M. et al. *DNA and Cell Biology* (1995) 14(1): 25; *see* Figure 1 for reported nucleic acid and amino acid sequences for GPR6). GPR4 has also been identified as an orphan GPCR (Heiber, M. et al, 14 *DNA Cell Biol.* 25 (1995)). OGR1, an orphan GPCR, is reported to have a high level of homology with GPR4 (Xu, Y. and Casey, G., 35 *Genomics* 397 (1996)). GPR21 is a 349 amino acid G protein coupled receptor for which the endogenous ligand is unknown (*see* GenBank Accession # U66580 for nucleic acid and deduced amino acid sequence). GPR21 has been reported to be located at chromosome 9q33. O'Dowd B. et al., 187 *Gene* 75 (1997). AL022171 is a human DNA sequence from clone 384F21 on chromosome 1q24. AL022171 has been identified to contain an open reading frame of 1,086 bp encoding for a 361 amino acid protein. (*see* GenBank Accession number AL022171). AL022171 is 68% homologous to GPR21 (*see* Figure 5B). GHSR is also identified as an orphan GPCR (Howard, A.D. et al, 273 *Science* 974 (1996)).

SUMMARY OF THE INVENTION

Disclosed herein are methods for screening of candidate compounds against endogenous, constitutively activated G protein-coupled orphan receptors (GPCRs) for the direct identification of candidate compounds as agonists, inverse agonists or partial agonists

to such receptors. For such screening purposes, it is preferred that an endogenous, constitutively activated orphan GPCR:G protein – fusion protein be utilized.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows a generalized structure of a G protein-coupled receptor with the numbers assigned to the transmembrane helixes, the intracellular loops, and the extracellular loops.

Figure 2 schematically shows the two states, active and inactive, for a typical G protein coupled receptor and the linkage of the active state to the second messenger transduction pathway.

Figure 3 is computerized representation of a "dot-blot" showing the distribution of the orphan receptor GPR4 across a variety of human tissues (*see* Appendix A for grid-code).

Figure 4 is a diagram showing enhanced binding of [³⁵S]GTPγS to membranes prepared from 293T cells transfected with the orphan receptor GPR3 compared to those transfected with control vector alone at 75 μg/well membrane protein. The radiolabeled concentration of [³⁵S]GTPγS was held constant at 1.2 nM and the GDP concentration was held constant at 1 μM. The assay was performed on 96-well format in Wallac scintistrips.

Figure 5A shows the amino acid alignment of orphan receptors GPR3, GPR6, and GPR12. **Figure 5B** shows the amino acid alignment of orphan receptors GPR21 and AL022171 (Consensus #1 indicates matching residues).

Figure 6A is a diagram showing that the orphan receptors GPR3, GPR6, and GPR 12 are confirmed to be constitutively active by their enhanced ability to induce expression of β-galactosidase from a CRE driven reporter system in VIP cells. **Figure 6B** and **6C** are diagrams of orphan receptors GPR21 and AL022171, respectively, that have also been confirmed to be

constitutively active by their enhanced ability to induce expression of the luciferase gene from a CRE driven reporter system in both 293 and 293T cells.

Figures 7A, 7B and 7C show the relative distribution of the expression of the GPR3 (A), GPR6 (B), and GPR12 (C) orphan receptors across several normal human tissues as determined by RT-PCR. Abbreviations: Ocx = occipital cortex; Hypoth = hypothalamus; Tex = temporal cortex; Fcx = frontal cortex.

Figures 8A and 8B show GPR3 receptor expression in normal (A) and epileptic (B) human brain tissue as examined by RT-PCR.

Figure 9A is a copy of an autoradiograph evidencing the results from in situ hybridization (normal rat) using GPR6 probe; **Figure 9B** is a reference map of the corresponding region of the rat brain.

Figure 10A is a copy of an autoradiograph evidencing the results from in situ hybridization (Zucker rat - lean) using GPR6 probe; **Figure 10B** is a copy of an autoradiograph evidencing the results from in situ hybridization (Zucker rat - obese) using GPR6 probe; **Figure 10C** is a reference map of the corresponding region of the rat brain.

Figures 11A-F are copies of autoradiographs evidencing the results from in situ hybridization (normal rat) using GPR12 probe.

Figure 12 is a copy of an autoradiograph evidencing the results from in situ hybridization (normal rat) using GPR6 probe (12A), and orexin 1 receptor probe (12B) with overlays for determination of co-localization of the two receptors (12C and 12D).

Figure 13 is a copy of an autoradiograph evidencing the results from in situ hybridization (normal rat) using GPR6 probe (13A), and melanocortin-3 receptor probe (13B) with overlays for determination of co-localization of the two receptors (13C and 13D).

Figure 14 provides results from co-localization experiment, evidencing that GPR6 and AGRP are co-localized within the arcuate. The arrow directs attention to to a specific cell within the arcuate, with the circle surrounding the cell; the "dots" are radiolabeled GPR6, and beneath those, in a darker shade, is AGRP.

Figure 15 provides graphic results of body weight over time from animals (n = 5) receiving antisense oligonucleotides to GPR6 (star symbol at Day 5 indicates day on which animals received d-amphetamine sulfate injection; *see* **Figure 16**).

Figure 16 provides bar graph results from baseline locomotor activity and from amphetamine-induced locomotive behavior in the animals of **Figure 15**.

Figure 17 provides bar-graph results from the direct identification of candidate compounds screened against GPR3 Fusion Protein (**Figure 17A**) and GPR6 Fusion Protein (**Figure 17B**).

Figure 18A-L is a sequence diagram of the preferred vector pCMV, including restriction enzyme site locations.

DETAILED DESCRIPTION

The scientific literature that has evolved around receptors has adopted a number of terms to refer to ligands having various effects on receptors. For clarity and consistency, the following definitions will be used throughout this patent document. To the extent that these definitions conflict with other definitions for these terms, the following definitions shall control:

AGONISTS shall mean materials (e.g., ligands, candidate compounds) that activate the intracellular response when they bind to the receptor, or enhance GTP binding to membranes.

AMINO ACID ABBREVIATIONS used herein are set out in Table 1:

TABLE 1

ALANINE	ALA	A
ARGININE	ARG	R
ASPARAGINE	ASN	N
ASPARTIC ACID	ASP	D
CYSTEINE	CYS	C
GLUTAMIC ACID	GLU	E
GLUTAMINE	GLN	Q
GLYCINE	GLY	G
HISTIDINE	HIS	H
ISOLEUCINE	ILE	I
LEUCINE	LEU	L
LYSINE	LYS	K
METHIONINE	MET	M
PHENYLALANINE	PHE	F
PROLINE	PRO	P
SERINE	SER	S
THREONINE	THR	T
TRYPTOPHAN	TRP	W
TYROSINE	TYR	Y
VALINE	VAL	V

PARTIAL AGONISTS shall mean materials (*e.g.*, ligands, candidate compounds) which activate the intracellular response when they bind to the receptor to a lesser degree/extent than do agonists, or enhance GTP binding to membranes to a lesser degree/extent than do agonists

ANTAGONIST shall mean materials (*e.g.*, ligands, candidate compounds) that competitively bind to the receptor at the same site as the agonists but which do not activate the intracellular response initiated by the active form of the receptor, and can thereby inhibit the intracellular responses by agonists or partial agonists. **ANTAGONISTS** do not diminish the baseline intracellular response in the absence of an agonist or partial agonist.

CANDIDATE COMPOUND shall mean a molecule (for example, and not limitation, a chemical compound) which is amenable to a screening technique. Preferably, the phrase "candidate compound" does not include compounds which were publicly known to be compounds selected from the group consisting of inverse agonist, agonist or antagonist to a receptor, as previously determined by an indirect identification process ("indirectly identified compound"); more preferably, not including an indirectly identified compound which has previously been determined to have therapeutic efficacy in at least one mammal; and, most preferably, not including an indirectly identified compound which has previously been determined to have therapeutic utility in humans.

COMPOSITION means a material comprising at least one component; a "pharmaceutical composition" is an example of a composition.

COMPOUND EFFICACY shall mean a measurement of the ability of a compound to inhibit or stimulate receptor functionality, as opposed to receptor binding affinity. A most preferred means of detecting compound efficacy is via measurement of GTP (via [³⁵S]GTPγS) or cAMP, as further disclosed in the Example section of this patent document.

CONSTITUTIVELY ACTIVATED RECEPTOR (Constitutively Active Receptor) shall mean a receptor subject to constitutive receptor activation. A constitutively activated receptor can be endogenous or non-endogenous.

CONSTITUTIVE RECEPTOR ACTIVATION shall mean stabilization of a receptor in the active state by means other than binding of the receptor with its endogenous ligand or a chemical equivalent thereof.

CONTACT or **CONTACTING** shall mean bringing at least two moieties together, whether in an in vitro system or an in vivo system.

DIRECTLY IDENTIFYING or **DIRECTLY IDENTIFIED**, in relationship to the phrase "candidate compound", shall mean the screening of a candidate compound against a constitutively activated receptor, preferably a constitutively activated orphan receptor, and most preferably against a constitutively activated G protein-coupled cell surface orphan receptor, and assessing the compound efficacy of such compound. This phrase is, under no circumstances, to be interpreted or understood to be encompassed by or to encompass the phrase "indirectly identifying" or "indirectly identified."

ENDOGENOUS shall mean a material that a mammal naturally produces. **ENDOGENOUS** in reference to, for example and not limitation, the term "receptor," shall mean that which is naturally produced by a mammal (for example, and not limitation, a human) or a virus. By contrast, the term **NON-ENDOGENOUS** in this context shall mean that which is not naturally produced by a mammal (for example, and not limitation, a human) or a virus. For example, and not limitation, a receptor which is not constitutively active in its endogenous form, but when manipulated becomes constitutively active, is most preferably referred to herein as a "non-endogenous, constitutively activated receptor." Both terms can be utilized to describe both "in vivo" and "in vitro" systems. For example, and not limitation, in a screening approach, the endogenous or non-endogenous receptor may be in reference to an in vitro screening system. As a further example and not limitation, where the genome of a mammal has been manipulated to include a non-endogenous constitutively activated receptor, screening of a candidate compound by means of an in vivo system is viable.

G PROTEIN COUPLED RECEPTOR FUSION PROTEIN and **GPCR FUSION PROTEIN**, in the context of the invention disclosed herein, each mean a non-endogenous protein comprising an endogenous, constitutively activated orphan GPCR fused to at least one

G protein, most preferably, the alpha (α) subunit of such G protein (this being the subunit that binds GTP), with the G protein preferably being of the same type as the G protein that naturally couples with endogenous orphan GPCR. For example, and not limitation, in an endogenous state, the G protein "G α " is the predominate G protein that couples with GPR6 such that a GPCR Fusion Protein based upon GPR6 would be a non-endogenous protein comprising GPR6 fused to G α . The G protein can be fused directly to the c-terminus of the endogenous, constitutively active orphan GPCR, or there may be spacers between the two.

INDIRECTLY IDENTIFYING or **INDIRECTLY IDENTIFIED** means the traditional approach to the drug discovery process involving identification of an endogenous ligand specific for an endogenous receptor, screening of candidate compounds against the receptor for determination of those which interfere and/or compete with the ligand-receptor interaction, and assessing the efficacy of the compound for affecting at least one second messenger pathway associated with the activated receptor.

INHIBIT or **INHIBITING**, in relationship to the term "response" shall mean that a response is decreased or prevented in the presence of a compound as opposed to in the absence of the compound.

INVERSE AGONISTS shall mean materials (*e.g.*, ligand, candidate compound) which bind to either the endogenous form of the receptor or to the constitutively activated form of the receptor, and which inhibit the baseline intracellular response initiated by the active form of the receptor below the normal base level of activity which is observed in the absence of agonists or partial agonists, or decrease GTP binding to membranes. Preferably, the baseline intracellular response is inhibited in the presence of the inverse agonist by at least 30%, more

preferably by at least 50%, and most preferably by at least 75%, as compared with the baseline response in the absence of the inverse agonist.

LIGAND shall mean an endogenous, naturally occurring molecule specific for an endogenous, naturally occurring receptor.

ORPHAN RECEPTOR shall mean an endogenous receptor for which the endogenous ligand specific for that receptor has not been identified or is not known.

PHARMACEUTICAL COMPOSITION shall mean a composition comprising at least one active ingredient, whereby the composition is amenable to investigation for a specified, efficacious outcome in a mammal (for example, and not limitation, a human). Those of ordinary skill in the art will understand and appreciate the techniques appropriate for determining whether an active ingredient has a desired efficacious outcome is based upon the needs of the artisan.

NON-ORPHAN RECEPTOR shall mean an endogenous naturally occurring molecule specific for an endogenous naturally occurring ligand wherein the binding of a ligand to a receptor activates an intracellular signaling pathway.

STIMULATE or **STIMULATING**, in relationship to the term "response" shall mean that a response is increased in the presence of a compound as opposed to in the absence of the compound.

The order of the following sections is set forth for presentational efficiency and is not intended, nor should be construed, as a limitation on the disclosure or the claims to follow.

A. Introduction

The traditional study of receptors has always proceeded from the a priori assumption (historically based) that the endogenous ligand must first be identified before discovery could

proceed to find antagonists and other molecules that could affect the receptor. Even in cases where an antagonist might have been known first, the search immediately extended to looking for the endogenous ligand. This mode of thinking has persisted in receptor research even after the discovery of constitutively activated receptors. What has not been heretofore recognized is that it is the active state of the receptor that is most useful for discovering agonists, partial agonists, and inverse agonists of the receptor. For those diseases which result from an overly active receptor, what is desired in a therapeutic drug is a compound which acts to diminish the active state of a receptor, not necessarily a drug which is an antagonist to the endogenous ligand. This is because a compound (drug) which reduces the activity of the active receptor state need not bind at the same site as the endogenous ligand. Thus, as taught by a method of this invention, any search for therapeutic compounds should start by screening compounds against the ligand-independent active state. The search, then, is for an inverse agonist to the active state receptor.

Screening candidate compounds against the endogenous, constitutively activated orphan receptors, for example, and not limited to, the endogenous, constitutively active GPCRs set forth herein, GPR3, GPR4, GPR6, GPR12, GPR21, GHSR, OGR1, RE2 and AL022171, allows for the direct identification of candidate compounds which act at these orphan cell surface receptors, without requiring any prior knowledge or use of the receptor's endogenous ligand. By determining areas within the body where such receptors are expressed and/or over-expressed, it is possible to determine related disease/disorder states which are associated with the expression and/or over-expression of these receptors; such an approach is disclosed in this patent document.

B. Disease/Disorder Identification and/or Selection

As will be set forth in greater detail below, most preferably inverse agonists to endogenous, constitutively activated orphan receptors, *e.g.*, such as those set forth herein (GPR3, GPR4, GPR6, GPR12, GPR21, GHSR, OGR1, RE2 and AL022171) can be identified by the methodologies of this invention. Such inverse agonists are ideal candidates as lead compounds in drug discovery programs for treating diseases related to these receptors. Indeed, an antagonist to such a receptor (even if the ligand were known) may be ineffective given that the receptor is activated even in the absence of ligand-receptor binding. Because of the ability to directly identify inverse agonists to these receptors, thereby allowing for the development of pharmaceutical compositions, a search, for diseases and disorders associated with these receptors is possible. For example, scanning both diseased and normal tissue samples for the presence of these orphan receptors now becomes more than an academic exercise or one which might be pursued along the path of identifying an endogenous ligand. Tissue scans can be conducted across a broad range of healthy and diseased tissues. Such tissue scans provide a preferred first step in associating a specific receptor with a disease and/or a disorder.

Preferably, the DNA sequence of the endogenous, constitutively activated GPCR is used to make a probe for RT-PCR identification of the expression of the receptor in tissue samples. The presence of a receptor in a diseased tissue, or the presence of the receptor at elevated concentrations in diseased tissue compared to normal tissue, can be utilized to identify a correlation with that disease. Receptors can equally well be localized to regions of organs by this technique. Based on the known functions of the specific tissues to which the receptor is localized, the putative functional role of the receptor can be deduced.

C. Homology Identification

The identification and association of an orphan receptor with diseases and/or disorders can be beneficially enhanced via identification of additional receptors having homology with the original orphan receptor. This approach was utilized in the identification of both GPR6 and GPR12, based upon their sequence homology with GPR3, and in the identification of AL022171, having sequence homology to GPR21. GPR3 was previously identified as a constitutively activated orphan receptor (*see Eggerick, supra*). What was not known, prior to this invention, was that GPR6, GPR12, GPR21 and AL022171 are also constitutively active in their endogenous states. Using known computerized databases (*e.g.*, dbEST), GPR6, GPR12, GPR21 and AL022171 were identified.

This highlights certain unique benefits of the invention disclosed herein: because the dogma in drug screening relies upon knowledge and identification of a receptor's endogenous ligand, the art had no motivation to explore whether or not GPR3 homologs were constitutively active in their endogenous forms (other than for, at best, academic curiosity). However, with the power of the present invention to directly identify inverse agonists to such receptors, coupled with the ability to locate the distribution of such receptors in tissue samples, the present invention dramatically transcends such idle curiosity and provides a means for alleviating diseases and disorders which impact the human condition.

D. Screening of Candidate Compounds

1. Generic GPCR screening assay techniques

When a G protein receptor becomes constitutively active, it binds to a G protein (*eg.*, Gq, Gs, Gi, Go) and stimulates the binding of GTP to the G protein. The G protein then acts as a GTPase and slowly hydrolyzes the GTP to GDP, whereby the receptor, under normal conditions, becomes deactivated. However, constitutively activated receptors continue to exchange GDP to

GTP. A non-hydrolyzable analog of GTP, [³⁵S]GTPγS, can be used to monitor enhanced binding to membranes which express constitutively activated receptors. It is reported that [³⁵S]GTPγS can be used to monitor G protein coupling to membranes in the absence and presence of ligand. An example of this monitoring, among other examples well-known and available to those in the art, was reported by Traynor and Nahorski in 1995. The preferred use of this assay system is for initial screening of candidate compounds because the system is generically applicable to all G protein-coupled receptors regardless of the particular G protein that interacts with the intracellular domain of the receptor. It is in the context of the use of a GTP assay system that a GPCR Fusion Protein is preferably utilized.

B 2. *Specific GPCR screening assay techniques*

Once candidate compounds are identified using the "generic" G protein-coupled receptor assay (i.e. an assay to select compounds that are agonists, partial agonists, or inverse agonists), further screening to confirm that the compounds have interacted at the receptor site is preferred. For example, a compound identified by the "generic" assay may not bind to the receptor, but may instead merely "uncouple" the G protein from the intracellular domain. In the case of GPR3, GPR4, GPR6, GPR12, GPR21, GHSR, OGR1, RE2 and AL022171, it has been determined that these receptors couple the G protein Gs. Gs stimulates the enzyme adenylyl cyclase (Gi, on the other hand, inhibits this enzyme). Adenylyl cyclase catalyzes the conversion of ATP to cAMP; thus, because these receptors are activated in their endogenous forms, increased levels of cAMP are associated therewith (on the other hand, endogenously activated receptors which couple the Gi protein are associated with decreased levels of cAMP). *See, generally, "Indirect Mechanisms of Synaptic Transmission," Chpt. 8, From Neuron To Brain (3rd Ed.) Nichols, J.G. et al eds. Sinauer Associates, Inc. (1992).* Thus,

assays that detect cAMP can be utilized to determine if a candidate compound is an inverse agonist to the receptor (*i.e.*, such a compound which contacts the receptor would decrease the levels of cAMP relative to the uncontacted receptor). A variety of approaches known in the art for measuring cAMP can be utilized; a most preferred approach relies upon the use of anti-cAMP antibodies. Another type of assay that can be utilized is a whole cell second messenger reporter system assay. Promoters on genes drive the expression of the proteins that a particular gene encodes. Cyclic AMP drives gene expression by promoting the binding of a cAMP-responsive DNA binding protein or transcription factor (CREB) which then binds to the promoter at specific sites called cAMP response elements and drives the expression of the gene. Reporter systems can be constructed which have a promoter containing multiple cAMP response elements before the reporter gene, e.g., β -galactosidase or luciferase. Thus, an activated Gs receptor such as GPR3 causes the accumulation of cAMP which then activates the gene and expression of the reporter protein. The reporter protein such as β -galactosidase or luciferase can then be detected using standard biochemical assays (see, for example, Chen et al. 1995). A cAMP assay is particularly preferred.

The foregoing specific assay approach can, of course, be utilized to initially directly identify candidate compounds, rather than by using the generic assay approach. Such a selection is primarily a matter of choice of the artisan. With respect to GPR6, use of a modified, commercially available cAMP assay was initially utilized for the direct identification of inverse agonists.

C 3. *GPCR Fusion Protein*

The use of an endogenous, constitutively activated orphan GPCR for use in screening of candidate compounds for the direct identification of inverse agonists, agonists and partial agonists

provides a unique challenge in that, by definition, the endogenous receptor is active even in the absence of an endogenous ligand bound thereto. Thus, in order to differentiate between, *e.g.*, the endogenous receptor in the presence of a candidate compound and the endogenous receptor in the absence of that compound, with an aim of such a differentiation to allow for an understanding as to whether such compound may be an inverse agonist, agonist, partial agonist or have no affect on such a receptor, it is preferred that an approach be utilized that can enhance such differentiation. A preferred approach is the use of a GPCR Fusion Protein.

Generally, once it is determined that an endogenous orphan GPCR is constitutively active, using the assay techniques set forth above (as well as others), it is possible to determine the predominant G protein that couples with the endogenous GPCR. Coupling of the G protein to the GPCR provides a signaling pathway that can be assessed. Because it is most preferred that screening take place by use of a mammalian expression system, such a system will be expected to have endogenous G protein therein. Thus, by definition, in such a system, the endogenous, constitutively active orphan GPCR will continuously signal. In this regard, it is preferred that this signal be enhanced such that in the presence of, *e.g.*, an inverse agonist to the receptor, it is more likely that one will be able to more readily differentiate, particularly in the context of screening, between the receptor when it is or is not contacted with the inverse agonist.

The GPCR Fusion Protein is intended to enhance the efficacy of G protein coupling with the endogenous GPCR. The GPCR Fusion Protein appears to be important for screening with an endogenous, constitutively activated GPCR because such an approach increases the signal that is most preferably utilized in such screening techniques. Facilitating a significant "signal to noise" ratio is important for the screening of candidate compounds as disclosed herein.

The construction of a construct useful for expression of a GPCR Fusion Protein is within the purview of those having ordinary skill in the art. Commercially available expression vectors and systems offer a variety of approaches that can fit the particular needs of an investigator. One important criterion for such a GPCR Fusion Protein construct is that the endogenous GPCR sequence and the G protein sequence both be in-frame (preferably, the sequence for the endogenous GPCR is upstream of the G protein sequence) and that the "stop" codon of the GPCR must be deleted or replaced such that upon expression of the GPCR, the G protein can also be expressed. The GPCR can be linked directly to the G protein, or there can be spacer residues between the two (preferably no more than about 12, although this number can be readily ascertained by one of ordinary skill in the art). We have evaluated both approaches, and in terms of measurement of the activity of the GPCR, the results are substantially the same; however, there is a preference (based upon convenience) of use of a spacer in that some restriction sites that are not used will, effectively, upon expression, become a spacer. Most preferably, the G protein that couples to the endogenous GPCR will have been identified prior to the creation of the GPCR Fusion Protein construct. Because there are only a few G proteins that have been identified, it is preferred that a construct comprising the sequence of the G protein (*i.e.*, a universal G protein construct) be available for insertion of an endogenous GPCR sequence therein; this provides for efficiency in the context of large-scale screening of a variety of different endogenous GPCRs having different sequences.

E. Medicinal Chemistry

Generally, but not always, direct identification of candidate compounds is preferably conducted in conjunction with compounds generated via combinatorial chemistry techniques, whereby thousands of compounds are randomly prepared for such analysis. Generally, the

results of such screening will be compounds having unique core structures; thereafter, these compounds are preferably subjected to additional chemical modification around a preferred core structure(s) to further enhance the medicinal properties thereof. In this way, inverse agonists, agonists and/or partial agonists that are directly identified can be beneficially improved upon prior to development of pharmaceutical compositions comprising such compounds. Generally, it is preferred that the binding affinity of a directly identified compound selected for further refinement into a pharmaceutical composition have a binding affinity for the receptor of less than 100nM, although this is generally a preference selection based upon the particular needs of the artisan. Such techniques are known to those in the art and will not be addressed in detail in this patent document.

F. Pharmaceutical Compositions

Candidate compounds selected for further development can be formulated into pharmaceutical compositions using techniques well known to those in the art. Suitable pharmaceutically-acceptable carriers are available to those in the art; for example, see Remington's Pharmaceutical Sciences, 16th Edition, 1980, Mack Publishing Co., (Oslo et al., eds.).

EXAMPLES

The following examples are presented for purposes of elucidation, and not limitation, of the present invention. While specific nucleic acid and amino acid sequences are disclosed herein, those of ordinary skill in the art are credited with the ability to make minor modifications to these sequences while achieving the same or substantially similar results reported below. It is intended that equivalent, endogenous, constitutively activated human orphan receptor sequences having eighty-five percent (85%) homology, more preferably

having ninety percent (90%) homology, and most preferably having greater than ninety-five percent (95%) homology to GPR3, GPR4, GPR6, GPR12, GPR21, GHSR, OGR1, RE2 and AL022171 fall within the scope of any claims appended hereto.

Example 1

Preparation of In Situ Probes

In situ probes for GPR3, GPR6, and GPR12 were prepared. The following PCR protocol was utilized for all three probes: the reaction condition utilized was 1X rTth DNA polymerase buffer II, 1.5 mM Mg(OAc)₂, 0.2 mM each of the 4 nucleotides, 0.228 µg rat genomic DNA, 0.25 µM of each primer (see below) and 1 unit of rTth DNA polymerase (Perkin Elmer) in 50 µl reaction volume. The cycle condition was 30 cycles of 94°C for 1 min, 55°C for 1 min and 72°C for 45 sec with a Perkin Elmer Cetus 2400 thermal cycler.

1. *Rat GPR3 in situ probe*

Because the full length cDNA sequence for rat GPR3 is not data-base available, the DNA fragment for the in situ probe was obtained by PCR using a 3' degenerate oligonucleotide based on the published human and mouse GPR3 sequences in the middle of the transmembrane domain 3, and a 5' degenerate oligonucleotide near the beginning of the 5' extracellular domain. The sequences of the oligonucleotides utilized were as follows:

5'-GGAGGATCCATGGCCTGGTTCTCAGC-3' (SEQ.ID.NO.:1; 5' oligo)

5'-CACAAGCTTAGRCCRTCC MG RCA RTTCCA-3' (SEQ.ID.NO.: 2; 3' oligo)

where R=A or G, and M=A or C.

A 537 bp PCR fragment containing nucleotide 24 through to the middle of transmembrane 3 was digested with Bam HI and Hind III and was subcloned into a Bam HI-Hind III site of pBluescript.

2. *Rat GPR 6 in situ probe*

The in situ probe DNA fragment of rat GPR6 was obtained by PCR based on the published rat GPR6 cDNA sequences. The sequences of the oligonucleotides utilized were as follows:

5'-GGAGAAGCTTCTGGCGGCGATGAACGCTAG-3' (SEQ.ID.NO.: 3; 5' oligo)

5'-ACAGGATCCAGGTGGCTGCTAGCAAGAG-3' (SEQ.ID.NO.: 4; 3' oligo)

A 608 bp PCR fragment containing nucleotide -10 through to the middle of transmembrane domain 4 was digested with Bam HI and Hind III and was subcloned into Bam HI-Hind III site of pBluescript.

3. *Rat GPR12 in situ probe*

The in situ probe DNA fragment of rat GPR12 was obtained by PCR based on the published rat GPR12 cDNA sequences. The sequences of the oligonucleotides utilized were as follows:

5'-CTTAAGCTTAAAATGAACGAAGACCCGAAG-3' (SEQ.ID.NO.: 5; 5' oligo)

5'-GGAGGATCCCCAGAGCATCACTAGCAT-3' (SEQ.ID.NO.: 6; 3' oligo)

A 516 bp PCR fragment containing nucleotide -5 through to the middle of transmembrane domain 4 was digested with Bam HI and Hind III and subcloned into a Bam HI-Hind III site of pBluescript.

In situ probe sequences generated were as follows:

Rat GPR3 probe:

GGAGGATCCATGGCCTGGTTCTCAGCCGGCTCAGGCAGTGTGAATGTGAGCAT
AGACCCAGCAGAGGAACCTACAGGCCAGCTACACTGCTGCCCTCTCCCAGGG
CCTGGGATGTGGTGCTGTGCATCTCAGGCACCCTGGTGTCTGCGAGAATGCT
CTGGTGATGGCCATCATTGTGGGCACGCCTGCCTTCCGCGCCCCCATGTTCTG
CTGGTGGGCAGCTTGGCCGTAGCAGACCTGCTGGCAGGCCTGGGCCTGGTCCT
GCACTTCGCTGCTGACTTCTGTATTGGCTCACCAGAGATGAGCTTGGTGCTGGT
TGGCGTGCTAGCAACGGCCTTTACTGCCAGCATCGGCAGCCTGCTGGCCATCA
CCGTTGACCGCTACCTTTCCCTGTACAACGCCCTCACCTACTACTCAGAGACAA
CAGTAACTCGAACCTACGTGATGCTGGCCTTGGTGTGGGTGGGTGCCCTGGGC
CTGGGGCTGGTTCCCGTGCTGGCCTGGAAGTCCCGGGACGGTCTAAGCTT

(SEQ.ID.NO.: 7)

Rat GPR6 probe:

AAGCTTCTGGCGGCGATGAACGCTAGCGCCGCCGCGCTCAACGAGTCCCAGGTGGTGGCAGTAGCG
GCCGAGGGAGCGGCAGCTGCGGCTACAGCAGCAGGGACACCGGACACCAGCGAATGGGGACCTCCG
GCAGCATCCGCGGCGCTGGGAGGCGGCGGAGGACCTAACGGGTCACTGGAGCTGTCTTCGCAGCTG
CCCGCAGGACCCTCAGGACTTCTGCTTTCGGCAGTGAATCCCTGGGATGTGCTGCTGTGCGTGTCGGG
GACTGTGATCGCAGGCGAAAAATGCGCTGGTGGTGGCGCTCATCGCATCCACTCCCGCGCTGCGCACG
CCCATGTTTGTGCTCGTGGGTAGTCTGGCCACTGCTGACCTGCTGGCGGGCTGTGGCCTCATCCTACA
CTTCGTGTTCCAGTACGTGGTGCCCTCGGAGACTGTGAGCCTGCTCATGGTGGGCTTCCTGGTGGCGT
CCTTCGCCGCTCAGTCAGCAGCCTGCTCGCTATCACAGTGGACCGTTACCTGTCCCTTTACAACGCG

CTCACCTACTACTCGCGCCGGACCCTGTTGGGCGTGACCTCTTGCTAGCAGCCACCTGGATCC

(SEQ.ID.NO.: 8)

Rat GPR12 probe:

AAGCTTAAAATGAACGAAGACCCGAAGGTCAATTTAAGCGGGCTGCCTCGGGACTGTATAGAAGCT
GGTACTCCGGAGAACATCTCAGCCGCTGTCCCCTCCCAGGGCTCTGTTGTGGAGTCAGAACCCGAGC
TCGTTGTCAACCCCTGGGACATTGTCTTGTGCAGCTCAGGAACCCTCATCTGCTGTGAAAATGCCGTC
GTGGTCCTTATCATCTTCCACAGCCCCAGCCTGCGAGCACCCATGTTCTGCTGATAGGCAGCCTGGC
TCTTGCAGACCTGCTGGCTGGTCTGGGACTCATCATCAATTTTGTTTTTGCCTACCTGCTTCAGTCAGA
AGCCACCAAGCTGGTCACAATTGGACTCATTGTCGCCTCTTTCTCTGCCTCTGTCTGCAGTTTGCTGG
CTATCACTGTGGACCGCTACCTCTCGCTGTATTACGCCCTGACGTACCACTCCGAGAGGACCGTCACC
TTTACCTATGTCATGCTAGTGATGCTCTGGGGATCC (SEQ.ID.NO.: 9)

Example 2 Receptor Expression

1. *cDNA and Vectors*

With respect to GPR3 and GPR6, expression vectors comprising cDNA were generously supplied by Brian O'Dowd (University of Toronto). The vector for GPR3 cDNA was pcDNA3; the vector for GPR6 was pRcCMV (the coding region for GPR6 was subcloned into pCMV vector at a Hind III-Xba I site). GPR12 cDNA was prepared using the following protocol: Human GPR12 cDNA was obtained by PCR using human genomic DNA and a 5' primer from the 5' untranslated region with a Hind III restriction site, and a 3' primer from the 3' untranslated region containing a Bam HI site'. Primers had the following sequences: 5'-CTTAAGCTTGTGGCATTGTTGGTACT-3' (SEQ.ID.NO.: 10; 5' oligo)

5'-TCTGGATCCTTGGCCAGGCAGTGGAAGT-3 (SEQ.ID.NO.: 11; 3' oligo)

PCR was performed using rTth polymerase (Perkin Elmer) with the buffer system provided by the manufacturers, 0.25 μ M of each primer, 0.2 μ M of each of the four nucleotides and 0.2 μ g of genomic DNA as template . The cycle condition was 30 cycles of 94°C for 1 min, 57 °C for 1min and 72 °C for 1.5 min. The 1.2 kb PCR fragment was digested with Hind III and Bam HI, and subcloned into Hind III-Bam HI site of pCMV expression vector. The resulting cDNA clones were fully sequenced and consistent with published sequences.

With respect to GPR21, PCR was performed using genomic DNA as template and rTth polymerase (Perkin Elmer) with the buffer system provided by the manufacturer, 0.25 μ M of each primer, and 0.2 mM of each of the four nucleotides. The cycle condition was 30 cycles of 94°C for 1 min, 62°C for 1min and 72 °C for 1 min and 20 sec. The 5' PCR primer was kinased with the sequence:

5'-GAGAATTCACCTCCTGAGCTCAAGATGAACT-3' (SEQ.ID.NO.:12)

and the 3' primer contained a BamHI site with the sequence:

5'-CGGGATCCCCGTAAGTGAAGCACTTCAGAT-3' (SEQ.ID.NO.:13).

The resulting 1.1 kb PCR fragment was digested with BamHI and cloned into EcoRV-BamHI site of pCMV expression vector. Nucleic acid (SEQ.ID.NO.:14) and amino acid (SEQ.ID.NO.:15) sequences for human GPR21 were thereafter determined.

With respect to AL022171, PCR was performed using genomic DNA as template and rTth polymerase (Perkin Elmer) with the buffer system provided by the manufacturer, 0.25 μ M of each primer, and 0.2 mM of each of the four nucleotides. The cycle condition was 30 cycles of 94°C for 1 min, 54°C for 1min and 72 °C for 1 min and 20 sec. The 5' primer contains an HindIII site with the following sequence:

5'-AGGAAGCTTTAAATTTCCAAGCCATGAATG-3' (SEQ.ID.NO.:16)

and the 3' primer contained a EcoRI site with the following sequence:

5'-ACCGAATTCAGATTACATTTGATTACTATG-3' (SEQ.ID.NO.:17). The resulting 1.15 kb PCR fragment was digested with HindIII and EcoRI and cloned into HindIII-EcoRI site of pCMV expression vector. Nucleic acid (SEQ.ID.NO.:18) and amino acid (SEQ.ID.NO.:19) sequences for human AL022171 were thereafter determined and verified.

With respect to GPR4 (GenBank accession number L36148), expression vectors comprising the cDNA was generously supplied by Brian O'Dowd (University of Toronto). The vector for GPR4 cDNA was pcDNA3 and this subcloned into pCMV vector at a Hind III-XbaI site (the 5' untranslated region between HindIII and an ApaI site was trimmed by conducting digestion/self ligation).

With respect to RE2 (GenBank accession number AF091890), PCR was performed using human brain cDNA as template and rTth polymerase (Perkin Elmer) with the buffer system provided by the manufacturer, 0.25 μ M of each primer, and 0.2 mM of each of the four nucleotides. The cycle condition was 30 cycles of 94°C for 1 min, 62°C for 1min and 72 °C for 1 min and 30 sec. The 5' PCR primer contained an EcoRI site with the sequence

5' -AGCGAATTCTGCCCCACCCACGCCGAGGTGCT-3' (SEQ. ID. No. 20)

and the 3' primer contained a BamHI site with the sequence

5'-TGCGGATCCGCCAGCTCTTGAGCCTGCACA-3' (SEQ.ID.NO.:21). The 1.36 kb PCR fragment that resulted after two rounds of PCR was then digested with EcoRI and BamHI and cloned into EcoRI-BamHI site of pCMV. Nucleic acid (SEQ. ID. NO. 22) and amino acid sequence (SEQ. ID. NO. 23) was thereafter determined.

With respect to OGR1 (GenBank accession number U48405), PCR was performed using human genomic DNA as template and rTth polymerase (Perkin Elmer) with the buffer system provided by the manufacturer, 0.25 μ M of each primer, and 0.2 mM of each of the four nucleotides. The cycle condition was 30 cycles of 94°C for 1 min, 62°C for 1min and 72 °C for 1 min and 20 sec. The 5' PCR primer contained a HindIII site with the sequence 5'-GGAAGCTTCAGGCCCAAAGATGGGGAACAT-3' (SEQ. ID. No. 24) and the 3' primer contain a BamHI site with the sequence 5'-GTGGATCCACCCGCGGAGGACCCAGGCTAG-3' (SEQ.ID.NO.25). The resulting 1.14 kb PCR fragment was digested with HindIII and BamHI and cloned into HindIII-BamHI site pCMV. Nucleic acid (SEQ. ID. NO. 26) and amino acid sequence (SEQ. ID. NO. 27) was thereafter determined.

With respect to GHSR, PCR was performed using hippocampus cDNA as template and TaqPlus Precision polymerase (Stratagene) with the buffer system provided by the manufacturer, 0.25 μ M of each primer, and 0.2 mM of each 4 nucleotides. The cycle condition was 30 cycles of 94°C for 1 min, 68°C for 1min and 72°C for 1 min and 10 sec. For first round PCR, the 5' PCR primer sequence:

5'-ATGTGGAACGCGACGCCCAGCG-3' (SEQ.ID.NO.40)

and the 3' primer sequence:

5'-TCATGTATTAATACTAGATTCT-3' (SEQ.ID.NO.41).

Two microliters of the first round PCR was used as a template for the second round PCR where the 5' primer was kinased with sequence:

5'-TACCATGTGGAACGCGACGCCCAGCGAAGAGCCGGGGT-3' (SEQ.ID.NO.:42) and

the 3' primer contains an EcoRI site with the sequence:

5'-CGGAATTCATGTATTAATACTAGATTCTGTCCAGGCCCG-3' (SEQ.ID.NO.:43). The 1.1 kb PCR fragment was digested with EcoRI and cloned into blunt-EcoRI site of CMVp expression vector. Nucleic acid (SEQ.ID.NO.:44) and amino acid (SEQ.ID.NO.:45) sequences for human GHSR were thereafter determined.

2. *Transfection procedure*

On day one, 1×10^7 293 or 293T cells per 150mm plate were plated out. On day two, two reaction tubes were prepared (the proportions to follow for each tube are per plate): tube A was prepared by mixing between 8-20 μ g DNA (e.g., pCMV vector; pCMV vector with receptor cDNA; pCMV with GPCR Fusion Protein, *supra*) in 1-2ml serum free DMEM (Irvine Scientific, Irvine, CA); tube B was prepared by mixing 50-120 μ l lipofectamine (Gibco BRL) in 1-2ml serum free DMEM. Tubes A and B were then admixed by inversions (several times), followed by incubation at room temperature for 30-45min. The admixture is referred to as the "transfection mixture". Plated cells were washed with 1XPBS, followed by addition of 10-12ml serum free DMEM. 2.4ml of the transfection mixture was then added to the cells, followed by incubation for 4hrs at 37°C/5% CO₂. The transfection mixture was then removed by aspiration, followed by the addition of 25ml of DMEM/10% Fetal Bovine Serum. Cells were then incubated at 37°C/5% CO₂.

For GPCR Fusion Protein, preferred amounts to the above are as follows: 12 μ g DNA; 2ml serum free DMEM; 60 μ l lipofectamine; 293 cells 9 and an addition of 12ml serum free DMEM).

Example 3 **Tissue Distribution of GPCR**

For some orphan receptors, it will be apparent to those in the art that there is an understanding of the distribution of such receptors within, e.g., a human, or associated with a

disease state. However, for many orphan receptors, such information is not known, or will not be known. It is therefore preferred that some understanding of where such receptors may be distributed be understood; this allows for the ability to gain a predictive opportunity to associate a particular receptor with a disease state or disorder associated with the particular tissue where the receptor may be preferentially expressed. Using a commercially available mRNA dot-blot format, the distribution of endogenous, constitutively active GPCRs in various tissue types was assessed.

Preferably, the entire coding region of the receptor is used to generate a radiolabeled probe using a Prime-It II™ Random Primer Labeling Kit (Stratagene, #300385), according to the manufacturer's instructions. As an example, this approach was utilized for GPR4.

Human RNA Master Blot™ kit (Clontech, #7770-1) was hybridized with this probe and washed under stringent conditions, in accordance with manufacturer instructions. The blot was exposed to Kodak BioMax™ Autoradiography film overnight, at -80°C. Results are presented in Figure 3. Based upon these results, it is noted that GPR4 appears to be expressed throughout a variety of fetal tissue types (row G), as well as non-fetal heart (C1), and non-fetal lung (F1). This approach can be readily utilized for other receptors.

Example 4

GTP MEMBRANE BINDING SCINTILLATION PROXIMITY ASSAY

When a G protein-coupled receptor is in its active state, either as a result of ligand binding or constitutive activation, the receptor binds to a G protein (in the case of GPR3, GPR4, GPR6, GPR12, GPR21, GHSR, OGR1, RE2 and AL022171, Gs) and stimulates the binding of GTP to the G protein. The trimeric G protein-receptor complex acts as a GTPase and slowly hydrolyzes the GTP to GDP, at which point the receptor normally is deactivated. Constitutively activated receptors continue to exchange GDP for GTP. The non-hydrolyzable GTP analog, [³⁵S]GTPγS, can be utilized to demonstrate enhanced binding of [³⁵S]GTPγS to membranes expressing

constitutively activated receptors. The advantage of using [^{35}S]GTP γ S binding to measure constitutive activation is that: (a) it is generically applicable to all G protein-coupled receptors; (b) it is proximal at the membrane surface making it less likely to pick-up molecules which affect the intracellular cascade.

The assay utilizes the ability of G protein coupled receptors to stimulate [^{35}S]GTP γ S binding to membranes expressing the relevant receptors. The assay can, therefore, be used in the direct identification method to screen candidate compounds to known, orphan and constitutively activated G protein coupled receptors. The assay is generic and has application to drug discovery at all G protein coupled receptors.

The [^{35}S]GTP γ S assay was incubated in 20 mM HEPES, pH 7.4, binding buffer with 12 nM [^{35}S]GTP γ S and 75 μg membrane protein [e.g., 293T cells expressing GPR3] and 1 μM GDP for 1 hour. Wheatgerm agglutinin beads (25 μl ; Amersham) were then added and the mixture was incubated for another 30 minutes at room temperature. The tubes were then centrifuged at 1500 X g for 5 minutes at room temperature and then counted in a scintillation counter.

Referring to Figure 4, GPR3 receptor was determined to have increased activity as compared to control; this heightened activity is not the result of autocrine stimulation in that the data were obtained from membrane preparations, as opposed to whole cell preparations.

Example 5

Receptor Homology Determination

Following confirmation that GPR3 is a constitutively activated receptor, a homology search of the available G protein-coupled data banks (GeneBank), using the commercially available program, *DNA Star*, identified two highly homologous receptors, GPR6 and GPR12 (see Figure 5A); both of these receptors are orphan receptors. While the sequence of these receptors was previously "known" (i.e., they were available on the databases), it was not known that these

two receptors are constitutively activated in their endogenous forms (*see* Example 6, Figure 7). Furthermore, heretofore there would be no reason to search for such receptors for use in a drug discovery program in that the ligands therefore are not known or have not been identified. As such, the dogma approach to drug discovery would at best find the homology between GPR3, GPR6 and GPR12 of minor interest or, more likely, irrelevant.

Example 6
Analysis of Homologous Receptors For Constitutive Activation

Although a variety of cells are available to the art for the expression of proteins, it is most preferred that mammalian cells be utilized. The primary reason for this is predicated upon practicalities, *i.e.*, utilization of, *e.g.*, yeast cells for the expression of a GPCR, while possible, introduces into the protocol a non-mammalian cell which may not (indeed, in the case of yeast, does not) include the receptor-coupling, genetic-mechanism and secretory pathways that have evolved for mammalian systems – thus, results obtained in non-mammalian cells, while of potential use, are not as preferred as that obtained from mammalian cells. Of the mammalian cells, COS-7, 293 and 293T cells are particularly preferred, although the specific mammalian cell utilized can be predicated upon the particular needs of the artisan.

1. Analysis of GPR3, GPR6 and GPR12

To generate a β -galactosidase reporter containing multiple Gal4 binding sites, a Bgl II/ HindIII fragment was removed from the somatostatin promoter-containing plasmid 1.4(5xGal)CAT (Leonard, J. et al (1992) *PNAS USA* 89:6247-6251) and cloned into p β gal-Basic (Promega). The Bgl II/ HindIII fragment contains a variant of the minimal somatostatin promoter (from -71 bp to +50 bp relative to the transcription start site) in which the core 4bp of the cAMP Response Element (-46 to -43) were replaced with 5 copies of the recognition sequence for the yeast transcription factor Gal4. When this reporter is co-transfected with an expression plasmid

encoding a Gal4-CREB fusion protein, it is highly responsive to agents that increase the cAMP signaling pathway.

VIP2.0Zc is a cell line that has been stably transfected with the reporter gene β -galactosidase under the control of a cAMP responsive VIP promoter (Konig et al. Mol. Cell. Neuro. 1991, 2, 331-337). The cell line was used here to indirectly measure the accumulation of intracellular cAMP. Approximately 2 million cells were plated in 6 cm plate the day before transfection. DNA (5 μ g), for each receptor, was mixed with 2.5 ml serum-free DMEM containing 200 μ g/ml DEAE dextran and 100 μ M chloroquine, and added to a rinsed cell monolayer. After incubation for 90 min in a CO₂ incubator, the transfection medium was removed. The cells were washed with serum-free medium and supplemented with fresh complete medium. Twenty four hours after transfection, the cells were replated into 96-well plate at a density of 50 - 100 K per well and the β -galactosidase activity was assayed 48 to 72 hours after transfection.

The assay buffer contained 100 mM sodium phosphate, 2 mM MgSO₄, 0.1 mM MnCl₂, pH 8.0. The cells were washed with PBS, and 25 μ l /well of hypotonic lysis buffer consisting of 0.1 X assay buffer was added. Ten minutes later, 100 μ l of assay buffer containing 0.5% Triton X-100 and 40 mM β -mercaptoethanol was added to each well and incubation at room temperature continued for 10 minutes. The substrate solution containing 5 mg/ml chlorophenol red- β -D-galactopyranoside (CPRG) in assay buffer was added at 25 μ l/well and the plate was incubated at 37°C for 30 minutes before absorbance at 595 nm was measured with a plate reader.

GPR3, GPR6 and GPR12 were assayed using the foregoing system, and it was determined that both GPR6 and GPR12 are constitutively active. See Figure 6A.

2. Analysis of GPR21 and AL022171

293 and 293T cells were plated-out on 96 well plates at a density of 2×10^4 cells per well and were transfected, using Lipofectamine Reagent (BRL), the following day according to manufacturer instructions. A DNA/lipid mixture was prepared for each 6-well transfection as follows: 260ng of plasmid DNA in 100 μ l of DMEM were gently mixed with 2 μ l of lipid in 100 μ l of DMEM (the 260ng of plasmid DNA consisted of 200ng of a 8xCRE-Luc reporter plasmid, 50ng of pCMV comprising endogenous receptor or non-endogenous receptor or pCMV alone, and 10ng of a GPRS expression plasmid (GPRS in pcDNA3 (Invitrogen)). The 8XCRE-Luc reporter plasmid was prepared as follows: vector SRIF- β -gal was obtained by cloning the rat somatostatin promoter (-71/+51) at BglV-HindIII site in the p β gal-Basic Vector (Clontech). Eight (8) copies of cAMP response element were obtained by PCR from an adenovirus template AdpCF126CCRE8 (*see 7 Human Gene Therapy* 1883 (1996)) and cloned into the SRIF- β -gal vector at the Kpn-BglV site, resulting in the 8xCRE- β -gal reporter vector. The 8xCRE-Luc reporter plasmid was generated by replacing the beta-galactosidase gene in the 8xCRE- β -gal reporter vector with the luciferase gene obtained from the pGL3-basic vector (Promega) at the HindIII-BamHI site. Following 30 min. incubation at room temperature, the DNA/lipid mixture was diluted with 400 μ l of DMEM and 100 μ l of the diluted mixture was added to each well. 100 μ l of DMEM with 10% FCS were added to each well after a 4hr incubation in a cell culture incubator. The following day the transfected cells were changed with 200 μ l/well of DMEM with 10% FCS. Eight (8) hours later, the wells were changed to 100 μ l /well of DMEM without phenol red, after one wash with PBS. Luciferase activity were measured the next day using the LucLite™ reporter gene assay kit (Packard) following manufacturer instructions and read on a 1450 MicroBeta™ scintillation and luminescence counter (Wallac). Results are summarized in Figures 6B and 6C.

GPR21 and AL022171 were assayed using the foregoing system, and based upon these results, it was determined that both GPR21 and AL022171 are constitutively active in their endogenous forms. *See* Figure 6B and 6C.

3. Analysis of GPR4, RE2, OGR1 and GHSR

Using the protocols defined herein, GPR4, RE2, OGR1 and GHSR were analyzed and determined to be constitutively active in their endogenous forms (data not shown).

Example 7

Tissue Distribution of GPR3, GPR6 and GPR12

Tissue samples were examined for expression of these orphan receptors by comparative RT-PCR, using the following primers:

GPR3:

5'-CTGGTCCTGCACTTTGCTGC-3' (SEQ. ID. NO.: 28)

5'-AGCATCACATAGGTCCGTGTCAC-3' (SEQ.ID.NO.: 29)

These primers amplify a 194bp fragment.

GPR6:

5'-ACCAGAAAGGGTGTGGGTACACTG-3' (SEQ. ID. NO.: 30)

5'-GGAACGAAAGGGCACTTTGG-3' (SEQ. ID. NO.: 31)

These primers amplify a 249bp fragment.

GPR12:

5'-GCTGCCTCGGGATTATTTAG-3' (SEQ. ID. NO.: 32)

5'-GCCTATTAGCAGGAACATGGGTG-3' (SEQ. ID. NO.: 33)

These primers amplify a 220bp fragment.

These amplicons were designed to be non-overlapping, i.e., there is no sequence similarity between them, and to have similar T_m 's, such that each primer pair amplifies its respective target at the same optimal annealing temperature. This diminishes the chance that an amplicon from one primer pair will act as an annealing target for the other primers in the multiplex reaction, therefore reducing the chance of interference with other primer pairs.

Total RNA was extracted from tissue samples (human) using TRIzol™ Reagent (Gibco/BRL), following manufacturer instructions. cDNA was generated using 2mg total RNA and a First-Strand™ cDNA synthesis kit (Pharmacia). The cDNA samples were then diluted 1:3 in H₂O and comparative PCR was performed as described (Jensen, J. et al. (1996) J. Biol. Chem. 271:187490) in the presence of [³²P]dCTP. All reactions included the SP1-specific primers, which amplify a 300bp fragment, to serve as an internal control. Using the primers outlined above, under defined PCR conditions (1 cycle: 95°C, 5min; 23 cycles: 95°C, 30sec, 58°C, 30sec, 72°C, 1min; 1 cycle: 72°C, 10min) gave consistently reliable and quantitatively accurate results. It was further determined that the selected primer pairs did not interfere with each other when multiplexed. PCR products were visualized by denaturing gel electrophoresis (7M urea, 5% polyacrylamide (Long Ranger™ Solution, AT Biochemical, 0.6 XTBE) and subsequent autoradiography.

Figures 7A, 7B, and 7C show the distribution of GPR3, GPR6 and GPR12 across human tissues. This information allows for assessing disease states that are associated with such tissue, as well as determining specific regions within such tissue where such expression predominates, thus allowing for correlating such receptor expression with particular disease states. This, in turn, then allows for direct identification of compounds that impact such receptors, without the need to understand or know the endogenous ligand for such receptor. Further screening reveals that GPR3

is expressed in much higher levels in human epilepsy tissue samples (tissue source: temporal cortex), as compared with controls, as evidenced by RT-PCR analysis (Figure 8).

Example 8A

Functional Analysis – GPR6 (In Situ Analysis)

The distribution of GPR6 in the hypothalamus suggested possible involvement in feeding behavior. Accordingly, a functional analysis of this receptor was undertaken. In situ analysis was conducted as follows:

1. *Probe Design*

GPR6 probe was produced from a 450bp HindIII-ScaI fragment of the GPR6 receptor cloned into the HindIII-SmaI site of pBluescriptSK+. Riboprobes were produced using a T7 transcription system in a standard labeling reaction consisting of: 1µg of linearized plasmid, 2µl of 5x transcription buffer, 125µCi of ³⁵S-UTP, 150µM of GTP, CTP and ATP, 12.5mM dithiothreitol, 20U of RNase inhibitor and 6U of appropriate polymerase. The reaction was incubated at 37°C for 90 min., labeled probe being separated from free nucleotides over Sephadex G-50 spin columns.

2. *Tissue preparation*

Dissected tissue was frozen in isopentane cooled to -42°C and subsequently stored at -80 °C prior to sectioning on a cryostat maintained at -20°C. Slide-mounted tissue sections were then stored at -80 °C.

3. *In Situ Hybridization Protocol*

Tissue sections were removed from the -80°C freezer and incubated with a 1 µg/ml solution of proteinase-K to permeabilize the tissue and inactivate endogenous RNase. After this treatment, sections were incubated in succession in water (1 min), 0.1 M triethanolamine (pH 8.0; 1 min), and 0.25% acetic anhydride in 0.1 M triethanolamine (10 min). The tissue

was then washed in 2 x SSC (0.3 mM NaCl, 0.03 mM Na citrate, pH 7.2; 5 min) and dehydrated through graded concentrations of ethanol. Sections were then hybridized with 1.5×10^6 dpm of [^{35}S]UTP-labeled cRNA probes in 20 μl of a hybridization buffer containing 75% formamide, 10% dextran sulfate, 3 x SSC, 50 mM sodium phosphate buffer (pH 7.4), 1 x Denhart's solution, 0.1 mg/ml yeast tRNA, and 0.1 mg/ml sheared salmon sperm DNA. Tissue sections were covered with coverslips that were sealed with rubber cement. The slides were incubated overnight at 50°C. On the following day, the rubber cement was removed, the coverslips were soaked-off with 2 x SSC, and the tissue sections were washed for 10 min in fresh 2 x SSC solution. Single stranded probe not hybridized with endogenous mRNAs was removed by incubating the sections for 30 min in 200 $\mu\text{g/ml}$ solution of RNase-A at 37°C. The tissue was then washed in increasingly stringent SSC solutions (2, 1 and 0.5 x SSC; 10 min each), followed by a 1 hr wash in 0.5 x SSC at 60°C. After this final wash, the tissue sections were dehydrated using graded concentrations of ethanol, air dried and prepared for detection by x-ray autoradiography on Kodak XAR-5 film.

4. *Analysis*

Utilizing the above protocol on normal male rats (Sprague-Dawley, Charles River), it was determined that GPR6 is expressed in the following areas of the brain: hypothalamus, hippocampus, nucleus accumbens, caudate and cerebral cortex. See Figure 9A for a representative tissue section (GPR6 receptor is presented in the dark areas; Figure 9B provides a reference map of the rat brain.)

Given the high levels of expression of GPR6 in areas of the brain associated with feeding, an in situ analysis was conducted using the above protocol on both lean and obese male Zucker rats (Charles River). As those in the art appreciate, the Zucker animals are

genetically bred to result in animals that exhibit a lean or obese phenotype. Figure 10A provides a representative tissue section of GPR6 receptor expression in the lean Zucker animals; Figure 10B provides a representative tissue section of GPR6 receptor expression in the obese Zucker animals; Figure 10C is a reference map of this section of the rat brain. These results support the position that the endogenous, constitutively activated orphan receptor GPR6 is relatively overexpressed in a model of obesity.

Example 8B

Functional Analysis – GPR12 (In Situ Analysis)

In situ analysis for the GPR12 receptor was conducted as follows:

1. *Probe Design*

GPR12 probe was produced from a 515bp (NT5 – NT520) HindIII-BamHI fragment of the rat GPR12 receptor cloned into the HindIII-BamHI site of pBluescriptSK+. Riboprobes were produced using a T3/T7 transcription system in a standard labeling reaction consisting of: 1µg of linearized plasmid, 2µl of 5x transcription buffer, 125µCi of ³⁵S-UTP, 150µM of GTP, CTP and ATP, 12.5mM dithiothreitol, 20U of Rnase inhibitor and 6U of appropriate polymerase. The reaction was incubated at 37°C for 90 min., labeled probe being separated from free nucleotides over Sephadex G-50 spin columns.

2. *Tissue preparation*

Dissected tissue was frozen in isopentane cooled to -42°C and subsequently stored at -80 °C prior to sectioning on a cryostat maintained at -20°C. Slide-mounted tissue sections were then stored at -80°C.

3. *In Situ Hybridization Protocol*

Tissue sections were removed from the -80°C freezer and incubated with a 1 µg/ml solution of proteinase-K to permeabilize the tissue and inactivate endogenous RNase. After

this treatment, sections are incubated in succession in water (1 min), 0.1 M triethanolamine (pH 8.0; 1 min), and 0.25% acetic anhydride in 0.1 M triethanolamine (10 min). The tissue was then washed in 2 x SSC (0.3 mM NaCl, 0.03 mM Na citrate, pH 7.2; 5 min) and dehydrated through graded concentrations of ethanol. Sections were then hybridized with 1.5×10^6 dpm of [^{35}S]UTP-labeled cRNA probes in 20 μl of a hybridization buffer containing 75% formamide, 10% dextran sulfate, 3 x SSC, 50 mM sodium phosphate buffer (pH 7.4), 1 x Denhart's solution, 0.1 mg/ml yeast tRNA, and 0.1 mg/ml sheared salmon sperm DNA. Tissue sections were covered with coverslips that were sealed with rubber cement. The slides were incubated overnight at 50°C. On the following day, the rubber cement was removed, the coverslips were soaked-off with 2 x SSC, and the tissue sections were washed for 10 min in fresh 2 x SSC solution. Single stranded probe not hybridized with endogenous mRNAs was removed by incubating the sections for 30 min in 200 $\mu\text{g/ml}$ solution of RNase-A at 37°C. The tissue was then washed in increasingly stringent SSC solutions (2, 1 and 0.5 x SSC; 10 min each), followed by a 1 hr wash in 0.5 x SSC at 60°C. After this final wash, the tissue sections were dehydrated using graded concentrations of ethanol, air dried and prepared for detection by x-ray autoradiography on Kodak XAR-5 film.

4. *Analysis*

Utilizing the above protocol on normal male rats (Sprague-Dawley, Charles River), it was determined that GPR12 is expressed in the following areas of the brain: hippocampus (particularly in regions CA₃, CA₄ and the dentate gyrus; outer layers of the cerebral cortex; and the amygdala - all of these regions are well known in the art as associated with regions important for learning and memory); and thalamic relay nuclei, including the lateral geniculate nucleus, the medial geniculate nucleus and the lateral thalamic nucleus (regions related to

lateral relay functions, e.g., vision and hearing). *See* Figures 11A-F for representative tissue sections (GPR12 receptor is presented in the dark areas).

Example 8C

Functional Analysis – Co-Localization of GPR6 With Feeding-Behavior Receptors (In Situ Analysis)

The human orexin receptor OX₁R, previously an orphan GPCR (a.k.a., "HFGAN72"), has been localized in the lateral hypothalamic region of the brain and has been hypothesized to be involved in regulation of feeding behavior. Sakurai, T. et al 92(4) Cell 573 (1998). As noted in Sakurai, "pharmacological intervention directed at the orexin receptors may prove to be an attractive avenue toward the discovery of novel therapeutics for diseases involving dysregulation of energy homeostasis, such as obesity and diabetes mellitus." *Id* at 582. The melanocortin-3 receptor (MC-3) has also been identified, Gantz, I. Et al, 268(11) J. Biol. Chem. 8246 (1993), and is similarly associated with energy homeostasis.

An understanding of the neural pathways involved in the regulation and dysregulation of energy homeostasis is important for appreciation of hierarchical nuances that are critical for rational drug design. Merely affecting one receptor, particularly a receptor that is "downstream" of a more relevant receptor-pathway, may lead to a substantial expenditure of time and resources that ultimately results in the development of a pharmaceutical compound that may have little, if any, substantive impact on a particular disease state. For example, leptin, while clearly involved in some fashion with energy homeostasis, has not, to date, evidenced an opportunity for the development of a pharmaceutical product in the area of obesity. And while both the OX₁ and MC-3 receptors (as well as other melanocortin receptors) are also, in some manner, involved with energy homeostasis, development of pharmaceuticals based upon the traditional receptor "antagonist" approach may prove to be more frustrating than fruitful if, for example, these

receptors are not constitutively active in their endogenous forms, and, within the energy homeostasis pathway, there is a receptor that is constitutively active in its endogenous state. Indeed, the endogenous, constitutively active receptor would, by definition, continually signal whereas the endogenous, non-constitutively active receptors would require ligand-binding for such signaling. Thus, in the case of GPR6, which is not only constitutively active in its endogenous form, but also appears to be significantly up-regulated in an animal model of obesity, GPR6 would, in essence, "trump" other energy homeostasis related receptors in that even with complete blockage via receptor antagonists to these receptors, GPR6 would continue signaling. Thus, a determination of whether these receptors (and others within the energy homeostasis pathway) are co-localized within discrete, neuronal regions, is useful in providing a more refined receptor target for drug development.

In situ hybridization studies were performed as described above for GPR6, OX₁R and MC-3 receptors. For GPR6, the in situ probe utilized was as set forth in Example 7A. For OX₁R and MC-3, the probes were based upon the published rat sequences and were approximately 950bp and 441bp, respectively. Tissue preparation (normal rats) and in situ hybridization were substantially the same as set forth in Example 8A.

Results are presented in Figure 12 (GPR6 and OX₁R) and Figure 13 (GPR6 and MC-3), where a red filter was used for GPR6 hybridization and a green filter was used for OX₁R (Figure 12B) and MC-3 (Figure 13B). Figures 12C and 12D (a magnified version of 12A) are generated by overlay of Figures 12A and 12B; co-localization is evidenced by areas having an orange color (from the combination of red and green). Thus, in Figure 12C, it can be seen that GPR6 and OX₁R are co-localized in a sub-set of cells in the lateral arcuate and in the ventromedial hypothalamic nucleus, both of these regions being involved in the energy homeostasis pathways. A similar

overlay-procedure for Figures 13A (GPR6) and 13B (MC-3) provides evidence that these receptors are co-localized primarily in the lateral arcuate.

Information continues to develop within the art as to the neural pathways associated with feeding behavior. An important component of this pathway is the neuropeptide agouti-related peptide (AGRP) sometimes referred to as agouti-related transcript (ART). The expression of AGRP is largely restricted to the arcuate nucleus (*see* Flier, J. S. and Maratos-Flier, E., and Figure 1 therein). The cells that produce AGRP also produce neuropeptide Y (NPY). Animal studies have evidenced that administration of AGRP and administration of NPY leads to increases in feeding behavior and obesity. AGRP has also been shown to be an antagonist to the melanocortin 4 (MC-4) receptor, and antagonism of the MC-4 receptor is also known to increase feeding behavior an obesity. Thus, AGRP appears to be involved in at least two pathways associated with feeding behavior. As set forth below, it has been discovered that the GPR6 receptor is co-localized within cells that produce AGRP, and based upon the results set forth below in Example 8, coupled with the fact that GPR6 is an endogenous, constitutively activated GPCR, it is apparent that GPR6 is in some manner a potential "regulator" of the system – when expression of the GPR6 receptor is reduced via the use of antisense protocols (Example 9) there was a exceedingly rapid loss in body weight of the animals tested, suggesting that GPR6 may regulate the expression of AGRP.

Unlike the "overlay" approach above, the protocol set forth in Marks, D.L. et al, 3 *Mol. & Cell. Neuro.* 395 (1992) was utilized for assessment of co-localization. AGRP (the AGRP cRNA probe was synthesized from a 382bp fragment of AGRP cDNA cloned into Bluescript SK vector) was analyzed in conjunction with radiolabeled GPR6 and both were found to be co-localized in the arcuate (*see* Figure 14). Given the role that AGRP plays with respect to homeostasis, and further given hat GPR6 is constitutively active in its endogenous state, the results

obtained from Example 9, *infra*, would be consistent with these data in that the almost immediate, significant loss of weight can be understood in the context of GPR6 influencing AGRP.

Example 9

Functional Analysis – GPR6 (In Vivo Analysis: GPR6 Antisense)

Based upon the results developed from Example 7, and while not wishing to be bound by any particular theory, it was hypothesized that reduction in the expression of the GPR6 receptor would lead to a reduction in, *inter alia*, feeding behavior, metabolism, body weight, etc.; thus, by decreasing expression of this receptor via use of an antisense oligonucleotide, it was hypothesized that such animals would evidence changes in functional feeding behavior and/or feeding-related metabolism. Examination of this hypothesis was considered analogous to utilization of an inverse agonist to the receptor in that an inverse agonist would reduce the constitutive activity of the GPR6 receptor, akin to reducing the expression of the receptor itself. It is noted that such an approach results in "knock-down", as opposed to "knock-out", of the receptor, *i.e.*, in general, it is accepted that an antisense approach reduces expression of the target protein by approximately 30%.

Sixteen adult male Sprague-Dawley rats (Harlan, San Diego) were used for this study. Animals were vivarium-acclimated for at least one week prior to use. Animals were housed (groups of two) in hanging plastic cages with food and water available ad lib. Animals were weighed and handled for at least one day prior to surgery (to establish baseline weight) and throughout the study (to assess the effects of the treatment). Daily food intake for pairs of animals in a cage was assessed by weighing the food in the feeding trough each morning before and after refilling. Groups included antisense (n = 5), missense (n = 4) and sterile water (n = 5).

Surgeries were performed under sodium pentobarbital anesthesia (60 mg/kg), supplemented with halothane as necessary. Animals were stereotaxically implanted with a single cannula (brain infusion kit, Alza Pharmaceuticals) aimed at the lateral ventricle (bregma, AP -1.0, Lat -1.5, DV -3.8 from the surface of the brain). The inlet of the cannula was connected via flexible tubing to the outlet of an osmotic minipump (Model 2001, Alza Pharmaceuticals), that was implanted subcutaneously between the shoulder blades according to instructions provided by the manufacturer.

Pumps contained antisense oligonucleotide 5'-GsCTAGCGTTCATCGCCGsC-3' (SEQ.ID.NO.:34; antisense) (wherein the small "s" denotes a phosphorothioate linkage) or missense oligonucleotide 5'-CsTGGACTGTATCGCCCCsG-3' (SEQ.ID.NO.: 35; missense), or sterile water vehicle. Oligonucleotides were synthesized by Genset Corp and diluted to 2µg/µl in sterile water. Because the pumps utilized deliver 1µl/hour, animals received 48µg/day of antisense or missense oligonucleotides, or 24µl/day of sterile water. Pumps were primed prior to implant by incubation in sterile saline at 37°C for at least four hours prior to implant.

Five days after surgery, animals were treated with d-amphetamine sulfate; six days after surgery, baseline and amphetamine-stimulated locomotor behavior were examined; seven days after surgery, animals were euthanized and brains rapidly removed and frozen for histological analysis.

Animals were taken from the vivarium to the testing room, placed into an open field enclosure (*see* below), and baseline activity assessed for 30 minutes. At the end of 30 minutes, animals were briefly removed from the enclosure, injected with d-amphetamine sulfate (1.0 mg/kg s.c., diluted in sterile saline; National Institute on Drug Abuse Drug Supply Program), and immediately returned to the enclosure for 150 minutes. Locomotor behavior

was quantified at 10 minute intervals in order to follow the time-course of baseline and amphetamine-stimulated activity.

Baseline and amphetamine-stimulated locomotor behavior were assessed in a San Diego Instruments Flex Field System, consisting of 16" x 16" x 15" clear plexiglas open field enclosures. Photocell arrays (16 in each dimension) which surrounded the open fields were interfaced with a personal computer for collection of data. One array at 2" above the floor of an enclosure detected locomotor activity, and a second at 5" detected rearing behavior. The computer provided a variety of measures of locomotor activity, including total photocell beam breaks, time active, time resting, distance traveled, total number of rears, and time spent rearing (data not shown). During testing, the testing room was dimly lit by an overhead incandescent bulb, with white noise to mask outside sounds.

Results are presented in Figures 15 and 16. In Figure 15, it is noted that animals receiving the antisense oligonucleotide (GPR6 "knock-down" animals) had significantly greater loss of weight as compared with either the missense oligonucleotide-treated animals, or the control-treated animals. With respect to locomotor activity, the results of Figure 16 support the position that the base-line and amphetamine-treatment locomotor activities were substantially the same across all three groups.

Example 10

GPCR Fusion Protein Preparation

The design of the endogenous, constitutively activated GPCR-G protein fusion construct was accomplished as follows: both the 5' and 3' ends of the rat G protein Gs α (long form; Itoh, H. et al., 83 PNAS 3776 (1986)) were engineered to include a HindIII (5'-AAGCTT-3') sequence thereon. Following confirmation of the correct sequence (including the flanking HindIII sequences), the entire sequence was shuttled into pcDNA3.1(-)

(Invitrogen, cat. no. V795-20) by subcloning using the HindIII restriction site of that vector. The correct orientation for the Gs α sequence was determined after subcloning into pcDNA3.1(-). The modified pcDNA3.1(-) containing the rat Gs α gene at HindIII sequence was then verified; this vector was now available as a "universal" Gs α protein vector. The pcDNA3.1(-) vector contains a variety of well-known restriction sites upstream of the HindIII site, thus beneficially providing the ability to insert, upstream of the Gs protein, the coding sequence of an endogenous, constitutively active GPCR. This same approach can be utilized to create other "universal" G protein vectors, and, of course, other commercially available or proprietary vectors known to the artisan can be utilized – the important criteria is that the sequence for the GPCR be upstream and in-frame with that of the G protein.

Both GPR3-Gs α Fusion Protein construct and GPR6-Gs α Fusion Protein construct were then made as follows: primers were designed for both the GPR3 and GPR6. For GPR3, the primers were as follows:

5'-gatcTCTAGAATGATGTGGGGTGCAGGCAGCC-3' (SEQ. ID. NO. 36; sense)

5'-ctagGGTACCCGGACATCACTGGGGGAGCGGGATC-3' (SEQ. ID. NO. 37, antisense)

The sense and anti-sense primers included the restriction sites for XbaI and KpnI, respectively.

For GPR6, the primers were as follows:

5'-gatcTCTAGAATGCAGGGTGCAAATCCGGCC-3' (SEQ. ID. NO. 38, sense)

5'-ctagGGTACCCGGACCTCGCTGGGAGACCTGGAAC-3' (SEQ. ID. NO. 39, antisense).

The sense and anti-sense primers also contained restriction sites for XbaI and KpnI, respectively.

These restriction sites are available upstream of the HindIII site in the pcDNA3.1(-) vector.

PCR was then utilized to secure the respective receptor sequences for fusion within the Gs α universal vector disclosed above, using the following protocol for each: 100ng cDNA for

GPR3 and GPR6 was added to separate tubes containing 2ul of each primer (sense and anti-sense), 3uL of 10mM dNTPs, 10uL of 10XTaqPlus™ Precision buffer, 1uL of TaqPlus™ Precision polymerase (Stratagene: #600211), and 80uL of water. Reaction temperatures and cycle times for GPR3 were as follows: the initial denaturing step was done at 94°C for five minutes, and a cycle of 94°C for 30 seconds; 55°C for 30 seconds; 72°C for two minutes (repeated 30 times for GPR3). A final extension time was done at 72°C for ten minutes. For GPR6, the initial denaturing step was done at 96°C for seven minutes, and a cycle of 96°C for 30seconds, 55°C for 30 seconds, and 72°C for two minutes was repeated 30 times. A final extension time of ten minutes at 72°C was done for GPR6. Both PCR products for GPR3 and GPR6 were ran on a 1% agarose gel and then purified (data not shown). Each purified product was digested with XbaI and KpnI (New England Biolabs) and the desired inserts were isolated, purified and ligated into the Gs universal vector at the respective restriction site. The positive clones were isolated following transformation and determined by restriction enzyme digest; expression using 293 cells was accomplished following the protocol set forth *infra*. Each positive clone for GPR3:Gs – Fusion Protein and GPR6:Gs – Fusion Protein was sequenced and made available for the direct identification of candidate compounds.

GPCR Fusion Proteins were analyzed as above and verified to be constitutively active (data not shown).

Example 11

Protocol: Direct Identification of Inverse Agonists and Agonists Using [³⁵S]GTPγS

Although we have utilized endogenous, constitutively active GPCRs for the direct identification of candidate compounds as, *e.g.*, inverse agonists, for reasons that are not altogether understood, intra-assay variation can become exacerbated. Preferably, then, a GPCR Fusion Protein, as disclosed above, is utilized. We have determined that when such a protein is used,

intra-assay variation appears to be substantially stabilized, whereby an effective signal-to-noise ratio is obtained. This has the beneficial result of allowing for a more robust identification of candidate compounds.

It is important to note that the following results have been obtained using an *orphan* receptor; as that data support, it is possible, using the techniques disclosed herein, to directly identify candidate compounds that modulate the orphan receptor as inverse agonists, agonists and partial agonists, directly from a primary screen; indeed, the methods disclosed herein are sensitive enough to allow for direct identification of both inverse agonist and agonist modulators on the same assay plate.

1. Membrane Preparation

Membranes comprising the endogenous, constitutively active orphan GPCR fusion protein of interest (*see* Examples 2 and 10) and for use in the direct identification of candidate compounds as inverse agonists, agonists or partial agonists were prepared as follows:

(a) Materials

Membrane Scrape Buffer was comprised of 20mM HEPES and 10mM EDTA, pH 7.4; Membrane Wash Buffer was comprised of 20 mM HEPES and 0.1 mM EDTA, pH 7.4; Binding Buffer was comprised of 20mM HEPES, 100 mM NaCl, and 10 mM MgCl₂, pH 7.4

(b) Procedure

All materials were kept on ice throughout the procedure. Firstly, the media was aspirated from a confluent monolayer of cells, followed by rinse with 10ml cold PBS, followed by aspiration. Thereafter, 5ml of membrane Scrape Buffer was added to scrape cells; this was followed by transfer of cellular extract into 50ml centrifuge tubes (centrifuged at 20,000 rpm for 17 minutes at 4°C). Thereafter, the supernatant was aspirated and the pellet was resuspended in

30ml Membrane Wash Buffer followed by centrifuge at 20,000 rpm for 17 minutes at 4°C. The supernatant was then aspirated and the pellet resuspended in Binding Buffer. This was then homogenized using a Brinkman polytron™ homogenizer (15-20 second bursts until the all material was in suspension). This is referred to herein as "Membrane Protein".

2. Bradford Protein Assay

Following the homogenization, protein concentration of the membranes was determined using the Bradford Protein Assay (protein can be diluted to about 1.5mg/ml, aliquoted and frozen (-80°C) for later use; when frozen, protocol for use is as follows: on the day of the assay, frozen Membrane Protein is thawed at room temperature, followed by vortex and then homogenized with a polytron at about 12 x 1,000 rpm for about 5-10 seconds; it is noted that for multiple preparations, the homogenizer should be thoroughly cleaned between homogenization of different preparations).

(a) Materials

Binding Buffer (as per above); Bradford Dye Reagent; Bradford Protein Standard were utilized, following manufacturer instructions (Biorad, cat. no. 500-0006).

(b) Procedure

Duplicate tubes were prepared, one including the membrane, and one as a control "blank". Each contained 800ul Binding Buffer. Thereafter, 10ul of Bradford Protein Standard (1mg/ml) was added to each tube, and 10ul of membrane Protein was then added to just one tube (not the blank). Thereafter, 200ul of Bradford Dye Reagent was added to each tube, followed by vortex of each. After five (5) minutes, the tubes were re-vortexed and the

material therein was transferred to cuvettes. The cuvettes were then read using a CECIL 3041 spectrophotometer, at wavelength 595.

3. Direct Identification Assay

(a) Materials

GDP Buffer consisted of 37.5 ml Binding Buffer and 2mg GDP (Sigma, cat. no. G-7127), followed by a series of dilutions in Binding Buffer to obtain 0.2 uM GDP (final concentration of GDP in each well was 0.1 uM GDP); each well comprising a candidate compound, had a final volume of 200ul consisting of 100ul GDP Buffer (final concentration, 0.1uM GDP), 50ul Membrane Protein in Binding Buffer, and 50ul [³⁵S]GTPγS (0.6 nM) in Binding Buffer (2.5 ul [³⁵S]GTPγS per 10ml Binding Buffer).

(b) Procedure

Candidate compounds (Tripos, Inc., St. Louis, MO) were received in 96-well plates (these can be frozen at -80°C). Membrane Protein (or membranes with expression vector excluding the GPCR Fusion Protein, as control), were homogenized briefly until in suspension. Protein concentration was then determined using the Bradford Protein Assay set forth above. Membrane Protein (and control) was then diluted to 0.25mg/ml in Binding Buffer (final assay concentration, 12.5ug/well). Thereafter, 100 ul GDP Buffer was added to each well of a Wallac Scintistrip™ (Wallac). A 5ul pin-tool was then used to transfer 5 ul of a candidate compound into such well (*i.e.*, 5ul in total assay volume of 200 ul is a 1:40 ratio such that the final screening concentration of the candidate compound is 10uM). Again, to avoid contamination, after each transfer step the pin tool was rinsed in three reservoirs comprising water (1X), ethanol (1X) and water (2X) – excess liquid should be shaken from the tool after each rinse and dried with paper and kimwipes. Thereafter, 50 ul of Membrane Protein is added to each well (a control well comprising

membranes without the GPCR Fusion Protein is also utilized), and pre-incubated for 5-10 minutes at room temperature (the plates were covered with foil in that the candidate compounds obtained from Tripos are light sensitive). Thereafter, 50 μ l of [35 S]GTP γ S (0.6 nM) in Binding Buffer was added to each well, followed by incubation on a shaker for 60 minutes at room temperature (again, in this example, plates were covered with foil). The assay was then stopped by spinning of the plates at 4000 RPM for 15 minutes at 22°C. The plates were then aspirated with an 8 channel manifold and sealed with plate covers. The plates were then read on a Wallacc 1450 using setting "Prot. #37" (as per manufacturer instructions).

Exemplary results are presented in Figure 17A (GPR3:Gs Fusion Protein) and Figure 17B (GPR6:Gs Fusion Protein) where each designation is a well comprising a different candidate compound, standard deviations based upon the mean results of each plate are in dashed lines and the vertical lines are the percent response. Note in Figure 17A well designation C4 – this compound was directly identified as an inverse agonist to the GPR3 receptor. Note in Figure 17B wells designated G7 and H9 – these compounds were directly identified as an inverse agonist and a agonist, respectively, to the GPR6 receptor. In both cases, these are orphan receptors.

It is preferred that following such direct identification, IC₅₀ (inverse agonist) or EC₅₀ (agonist) values be determined; those having ordinary skill in the art are credited with utilizing IC₅₀ and EC₅₀ assay protocols of choice.

Example 12

Protocol: Confirmation Assay

After using an independent assay approach to provide a directly identified candidate compound as set forth above, it is preferred that a confirmation assay then be utilized. In this case, the preferred confirmation assay is a cyclase-based assay.

A modified Flash Plate™ Adenylyl Cyclase kit (New England Nuclear; Cat. No. SMP004A) was utilized for confirmation of candidate compounds directly identified as inverse agonists and agonists to endogenous, constitutively activated orphan GPCRs in accordance with the following protocol.

Transfected cells were harvested approximately three days after transfection. Membranes were prepared by homogenization of suspended cells in buffer containing 20mM HEPES, pH 7.4 and 10mM MgCl₂. Homogenization was performed on ice using a Brinkman Polytron™ for approximately 10 seconds. The resulting homogenate was centrifuged at 49,000 X g for 15 minutes at 4°C. The resulting pellet was then resuspended in buffer containing 20mM HEPES, pH 7.4 and 0.1 mM EDTA, homogenized for 10 seconds, followed by centrifugation at 49,000 X g for 15 minutes at 4°C. The resulting pellet can be stored at -80°C until utilized. On the day of direct identification screening, the membrane pellet is slowly thawed at room temperature, resuspended in buffer containing 20mM HEPES, pH 7.4 and 10mM MgCl₂, to yield a final protein concentration of 0.60mg/ml (the resuspended membranes are placed on ice until use).

cAMP standards and Detection Buffer (comprising 2 µCi of tracer [¹²⁵I cAMP (100 µl) to 11 ml Detection Buffer) were prepared and maintained in accordance with the manufacturer's instructions. Assay Buffer was prepared fresh for screening and contained 20mM HEPES, pH 7.4, 10mM MgCl₂, 20mM phosphocreatine (Sigma), 0.1 units/ml creatine phosphokinase (Sigma), 50 µM GTP (Sigma), and 0.2 mM ATP (Sigma); Assay Buffer can be stored on ice until utilized.

Candidate compounds identified as per above (if frozen, thawed at room temperature) were added to plate wells (3µl/well; 12µM final assay concentration), together with 40 µl Membrane Protein (30µg/well) and 50µl of Assay Buffer. This admixture was then incubated for 30 minutes at room temperature, with gentle shaking.

Following the incubation, 100 μ l of Detection Buffer was added to each well, followed by incubation for 2-24 hours. Plates were then counted in a Wallac MicroBeta™ plate reader using "Prot. #31" (as per manufacturer instructions).

Although a variety of expression vectors are available to those in the art, it is most preferred that the vector utilized be pCMV. This vector has been deposited with the American Type Culture Collection (ATCC) on October 13, 1998 (10801 University Blvd., Manassas, VA 20110-2209 USA) under the provisions of the Budapest Treaty for the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure. The vector was tested by the ATCC and determined to be viable. The ATCC has assigned the following deposit number to pCMV: ATCC #203351. A diagram of pCMV (including restriction sites) is set forth in Figure 18.

It is intended that each of the patents, applications, and printed publications mentioned in this patent document be hereby incorporated by reference in their entirety. As those skilled in the art will appreciate, numerous changes and modifications may be made to the preferred embodiments of the invention without departing from the spirit of the invention. It is intended that all such variations fall within the scope of the invention.

Appendix A

Figure 3 Grid Code

A2 – amygdala; A3 – caudate nucleus; A4 – cerebellum; A5 – cerebral cortex; A6 – frontal cortex;
A7 – hippocampus; A8 – medulla oblongata
B1- occipital cortex; B2 – putamen; B3 – substantia nigra; B4 – temporal cortex; B5 – thalamus;
B6 – sub-thalamic nucleus; B7 – spinal cord
C1 – heart; C2 – aorta; C3 – skeletal muscle; C4 – colon; C5 – bladder; C6 – uterus; C7 –
prostate; C8 – stomach
D1 – testis; D2 – ovary; D3 – pancreas; D4 – pituitary gland; D5 – adrenal gland; D6 – thyroid;
D7 – salivary gland; D8 – mammary gland
E1 – kidney; E2 – liver; E3 – small intestine; E4 – spleen; E5 – thymus; E6 – peripheral leukocyte;
E-8 lymph node; E9 – bone marrow
F1 – tonsil; F2 – lung; F3 – trachea; F4 – placenta
G1 – fetal brain; G2 – fetal heart; G3 – fetal kidney; G4 – fetal liver; G5 – fetal spleen; G6 – fetal
thymus; G8 – fetal lung